

# Scale-up of an RNA reference standard for high-throughput microarray QC

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(with thanks to a host of others  
at Agilent and Rosetta...)



**Agilent Technologies**

# Problem Statement

- **Goal & Constraints**

- + **Sample for final product lot QC (hybridization assay)**

- **Fixed QC array design**

- **Sensitivity to known & potential error modes**

- + **Sample must be**

- **Reproducible**

- **Manufacturable**

- **Economical**



# Possibilities Considered

- **Oligo-only sample**
- **Complex natural sample**
- **Complex synthetic sample**



# Possibilities Considered: Details

- **Oligo-only sample**
  - + **Easiest to make & maintain**
  - + **Limited relevance to customer experience**
    - **Narrow dynamic range**
    - **No cRNA component**
    - **Difficult to generate expression ratio data**
      - i. **Need to develop a family of oligos**
      - ii. **Need to develop QC and formulation methods to maintain ratios**



# Possibilities Considered: Details

- **Complex natural sample**

- + **Hardest to make and maintain**

- **More labor to make than to use**
    - **Qualifying new batches of natural mRNA difficult**
    - **Species-specific**

- + **Most relevant to customer experience**

- **Full dynamic range**
    - **Labeled cRNA, as used by customers**
    - **If 2 natural samples are used, they can be chosen to generate a rich range of differential expression (but TRUTH difficult to determine...)**

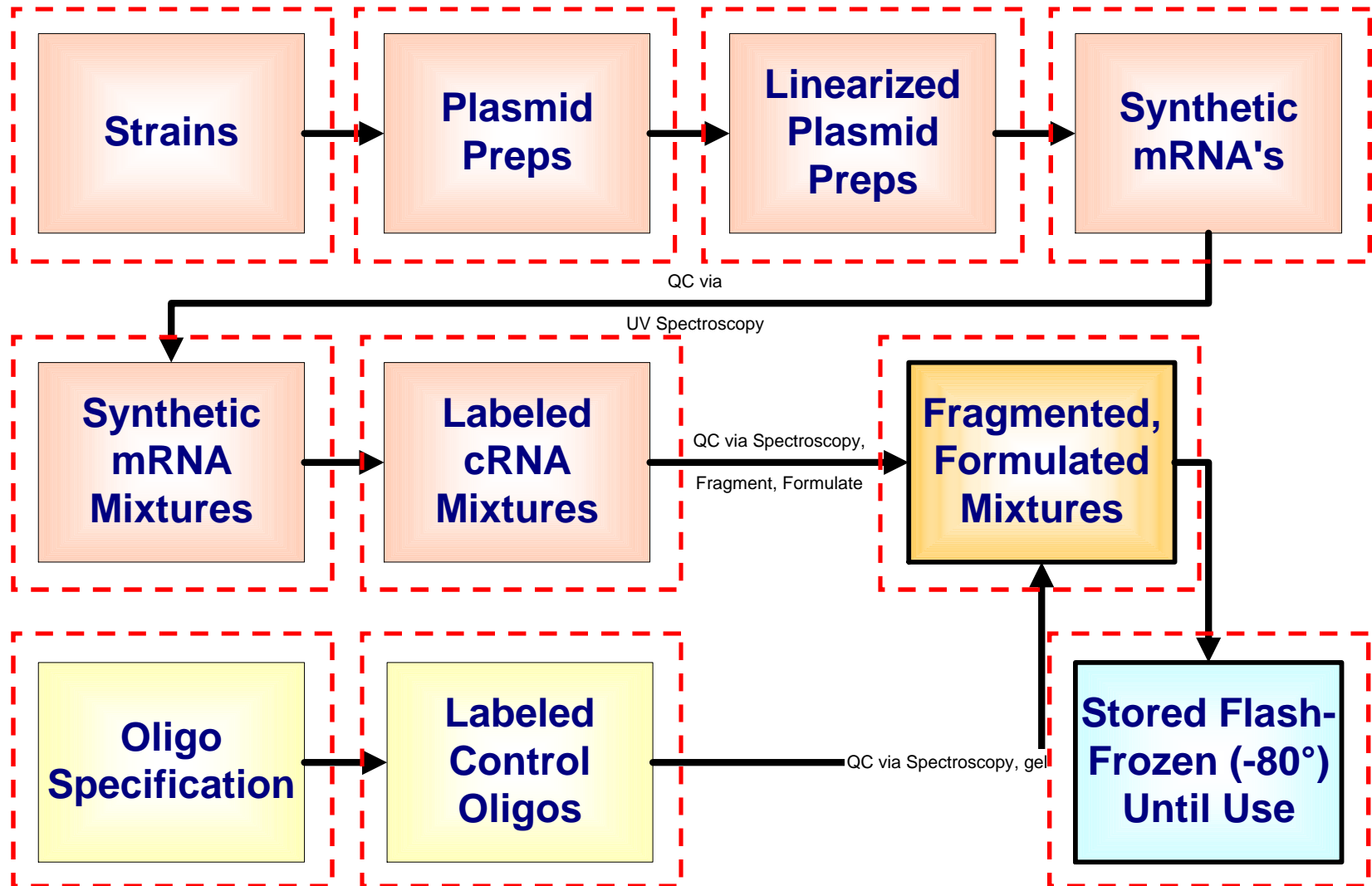


# Possibilities Considered: Details

- ✓ • **Complex synthetic sample (oligo + E1A cRNA)**
  - + Intermediate difficulty of production/maintenance**
    - 300x more efficient than “natural” sample
    - Stable source of (synthetic) mRNA component
    - Species-independent
  - + Intermediate relevance to customer experience**
    - ~200-fold dynamic range
    - Labeled cRNA, as used by customers
    - Known ratios of E1A targets in 2 samples
    - Control relative specific activities by labeling mixture



# Biomaterials Flow



# Typical E1A Cocktail Compositions\*

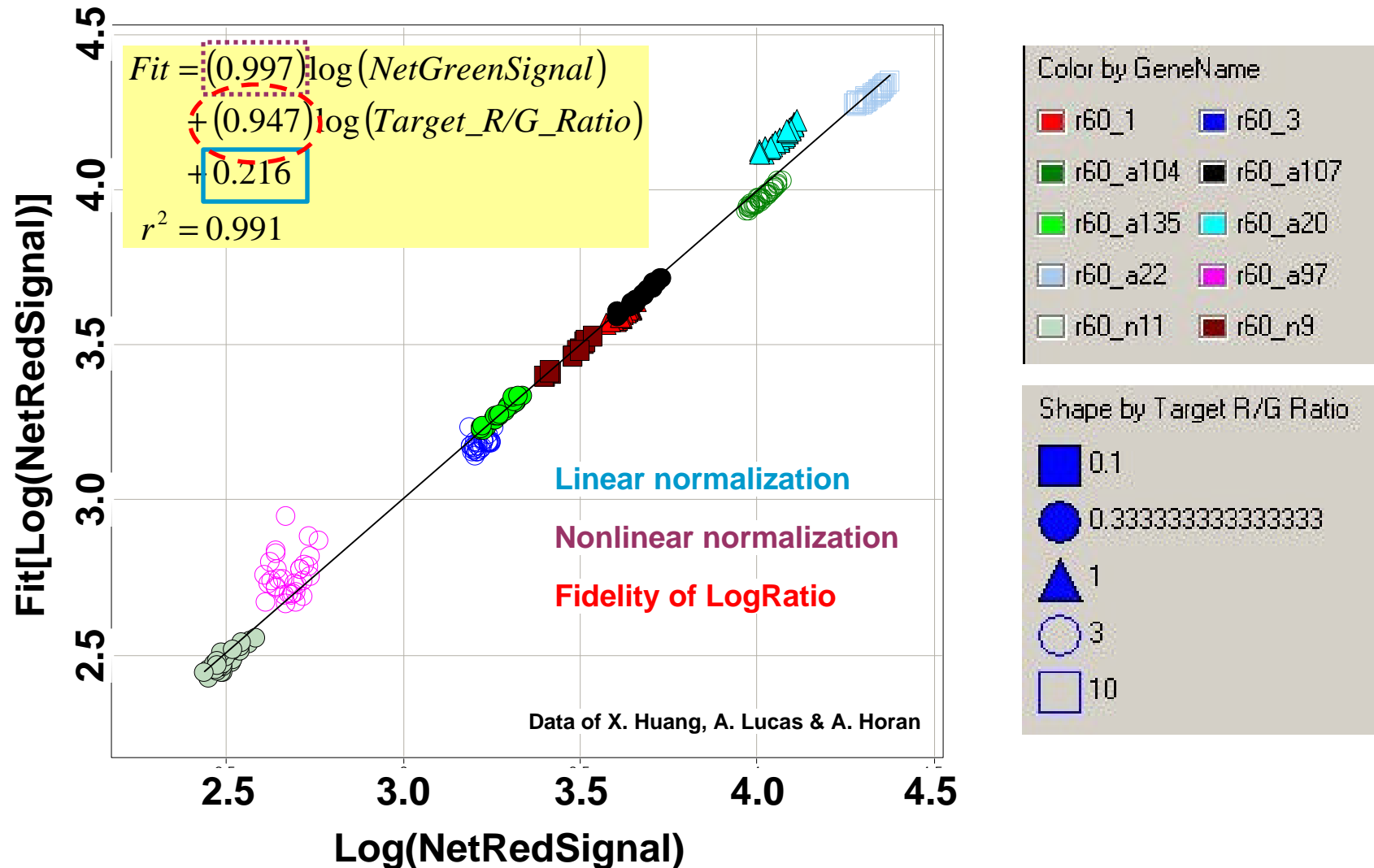
Transcript	Nominal Copies per Cell		Target Ratio
	Cocktail 11	Cocktail 12	
r60_a20	100	100	1:1
r60_1	10	10	1:1
r60_a22	10	100	1:10
r60_n9	100	10	10:1
r60_a104	10	30	1:3
r60_a107	30	10	3:1
r60_3	3	9	1:3
r60_a135	9	3	3:1
r60_a97	0.5	1.5	1:3
r60_n11	1.5	0.5	3:1

\* courtesy of Rosetta Inpharmatics





# Multiple Regression Model of E1A Data



# Fixed QC Array Design

**Probes to oligo targets:**

**Probes to cRNA targets:**

**Customer-oriented, ratio-centric measurements**

**Measurement of known printing and synthesis error modes**



# Routine Quality Metrics Based on E1A Targets

- **Ratio Sensitivity**

- + % of 1 copy/cell probes yielding ratios within 50% of expected value

- **Reproducibility**

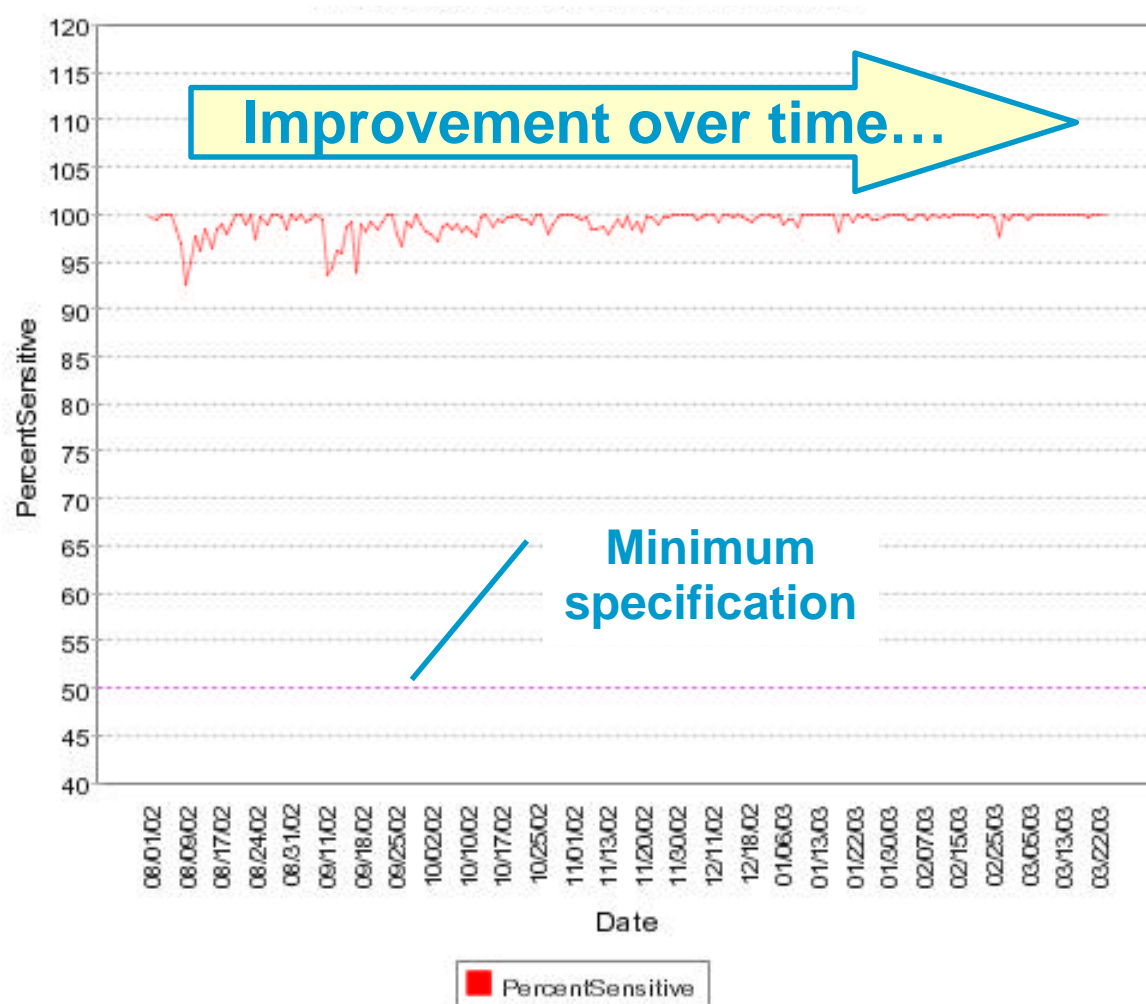
- + Maximum value of  $S_{\text{LogRatio}}$  for 5 different E1A probes
  - + Indicates CV of LogRatio Measurements

- **Accuracy**

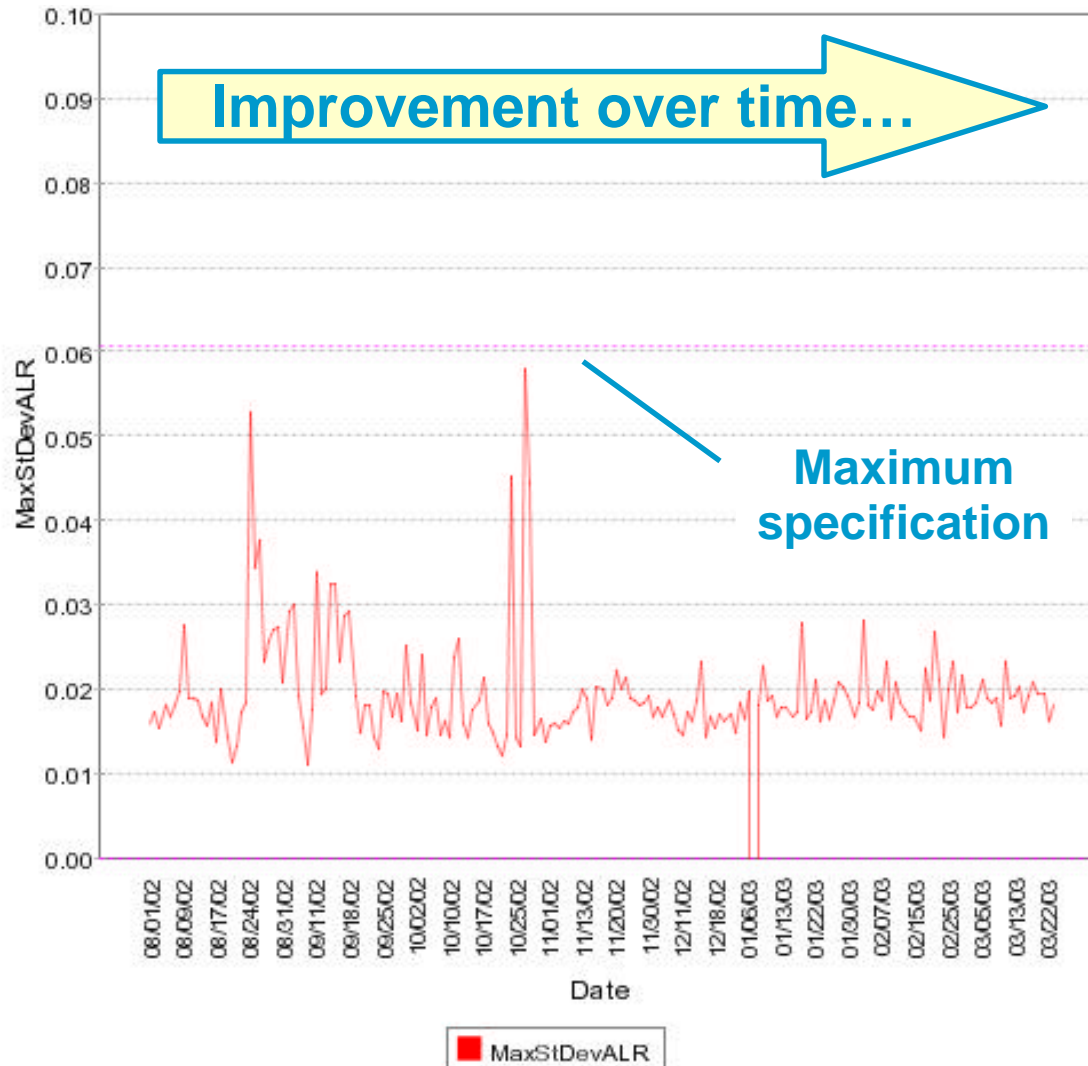
- + Average LogRatios (n=30) for 3:1 and 1:3 probes



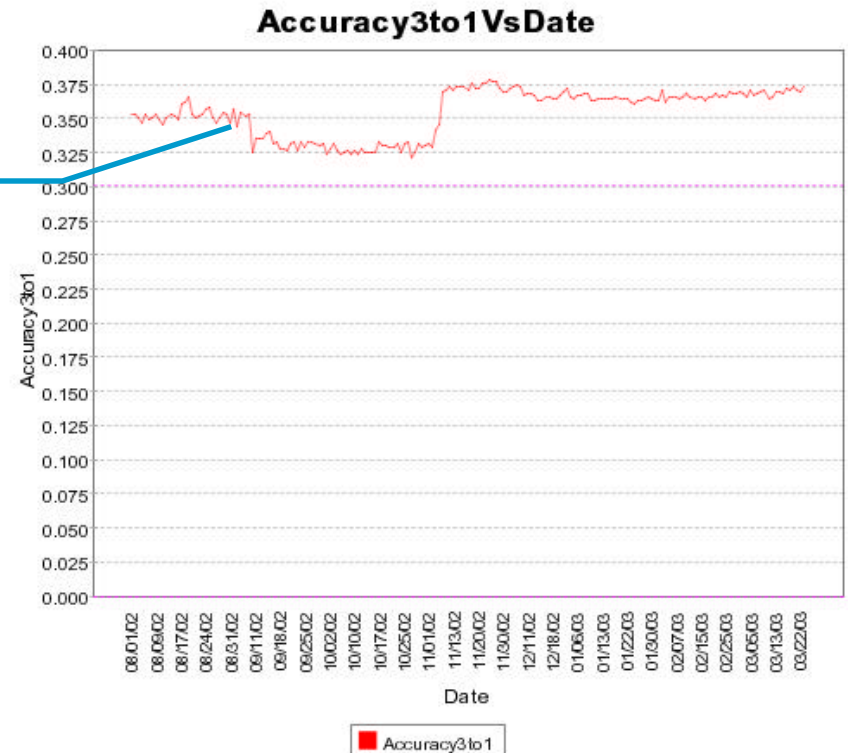
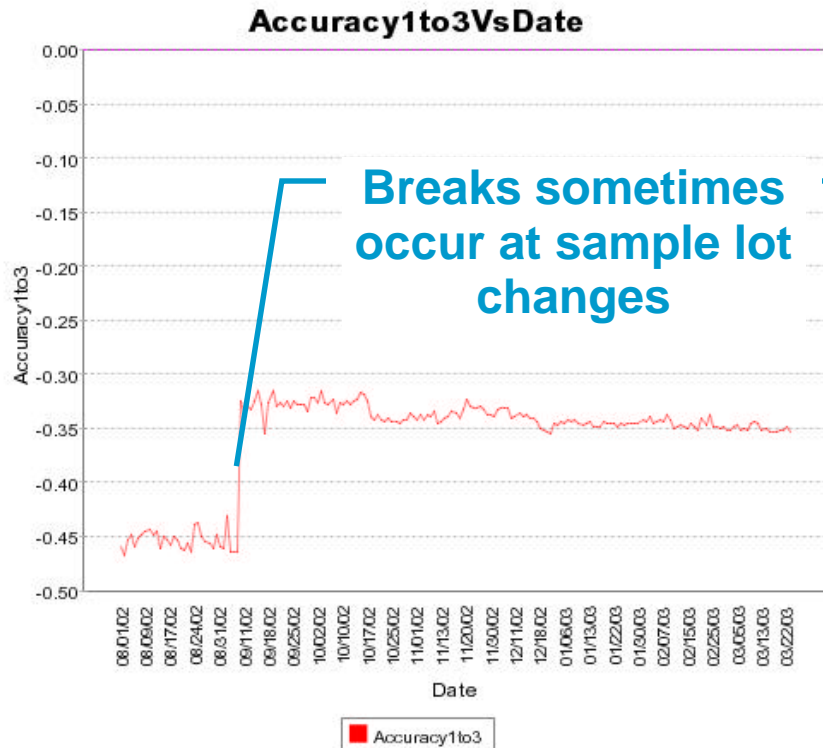
# Run Chart: Ratio Sensitivity Metric



# Run Chart: Reproducibility Metric



# Run Charts: Accuracy Metrics



# Conclusions

- **High-throughput microarray QC presents some unique challenges**
  - + **Long-term stability of standard sample**
  - + **Cost-effectiveness of standard sample**
- **The Rosetta E1A spike-in system can be used to perform high-throughput microarray QC**
  - + **Manufacturable (>10,000 assays to date)**
  - + **Good compromise between simple and complex samples**
  - + **Ratio-centric metrics**



# Relevance to NIST Microarray Standards Initiative

- **Synthetic mRNA samples are a viable approach to standard generation and maintenance**
- **Rosetta E1A standard set is particularly attractive**
  - + **Species-independent**
  - + **Proven track record**
  - + **Easily extended (by cloning additional inserts)**
  - + **Suitable for use in multiple systems & settings**

